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PREPARATION OF STEVIA EXTRACT USED IN THE FOOD INDUSTRY AND ITS ANTIMICROBIAL ACTIVITY

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A dry extract of the leaves of stevia, belonging to the Asteraceae family, grown in Uzbekistan, was extracted with hot water for use as a sweet flavor additive in the food industry. The content of the extract was determined by thin layer chromatography. The extract turned out to be a mixture of various steviol glycosides. It has been established that the main part of the mass of the extract is steviol glycosides - stevioside, rebaudioside A, rebaudioside C, rebaudioside B. When studying the biological activity of the resulting composition, its antimicrobial activity was determined.

Keywords: stevia, extract, sweet flavor additive, steviol glycosides, stevioside, rebaudioside, thin layer chromatography, antimicrobial activity

ПРИГОТОВЛЕНИЕ ЭКСТРАКТА СТЕВИИ, ИСПОЛЬЗУЕМОГО В ПИЩЕВОЙ ПРОМЫШЛЕННОСТИ, И ЕГО АНТИМИКРОБНАЯ АКТИВНОСТЬ

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Сухой экстракт листьев стевии, принадлежащей к семейству сложноцветных, выращиваемой в Узбекистане, экстрагировали горячей водой для использования в качестве сладкой вкусоароматической добавки в пищевой промышленности. Содержание экстракта определяли методом тонкослойной хроматографии. Экстракт оказался смесью различных гликозидов стевии. Установлено, что основную часть массы экстракта составляют гликозиды стевии - стевииозид, ребаудиозид А, ребаудиозид С, ребаудиозид В. При изучении биологической активности полученной композиции была определена ее антимикробная активность.

Ключевые слова: гидрогенизация, стевия, экстракт, сладкая вкусоароматическая добавка, стевииоловые гликозиды, стевииозид, ребаудиозид, тонкослойная хроматография, антимикробная активность

OZIQ-OVQAT SAN'OATIDA QO'LLANILADIGAN STEVIA EKSTRAKTINI OLISH VA UNING MIKROBLARGA QARSHI FAOLLIGI

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O'zbekistonda yetishtiriladigan oziq-ovqat san'otida shirin ta'm beruvchi qo'shimcha sifatida qo'llash uchun Asteraceae oilasiga mansub stevia o'simligining barglarini issiq suv yordamida ekstraksiya qilib quruq ekstrakti ajratib olindi. Ekstrakt tarkibini tahlil qilish uchun yupqa qatlamli xromatografiya usuli yordamida ekstrakt tarkibi tekshirildi. Ekstrakt turli xil steviol glikozidlari aralashmasidan iborat ekanligi aniqlandi. Ekstrakt tarkibidagi steviol glikozidlar - steviozid, rebaudiozid A, rebaudiozid C, rebaudiozid B massani asosiy qismini tashkil etishi aniqlandi. Olingan yig'ma tarkibi biologik faolligi o'rganilganda mikroblarga qarshi faolligi aniqlandi.

Kalit so'zlar: gidrogsteviya, ekstraktlar, shirin ta'm beruvchi qo'shimcha, steviol glikozidlar, steviozid, rebaudiozid, yupqa qatlamli xromatografiya, mikroblarga qarshi faollik

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Introduction

Stevia Rebaudiana Bertoni - is a perennial plant belonging to the Asteraceae family, which grows mainly in Paraguay and Brazil. Currently, the leaves of the *Stevia rebaudiana* plant have been used in the food industry since ancient times due to their sweet taste [1].

There are currently about 200 species of *Stevia* plants, but only one of them, *Stevia rebaudiana*,

contains steviol glycosides. The results of studies on steviol glycosides contained in *Stevia rebaudiana* showed that steviol glycoside - stevioside is found in all above-ground parts of the plant, but stevioside is 6-15% (relative to the obtained dry mass) in the leaves of the plant. There is very little in the stem [2]. Stevioside is registered in the food industry as a sweet flavoring E960 food additive.

Steviol glycosides are useful as sweeteners for people with impaired carbohydrate metabolism and especially for people with diabetes because they have hypoglycemic properties [3, 4].

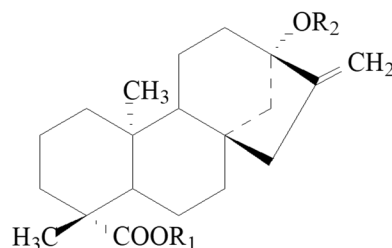
Stevia has been used in the food industry for over 40 years [5]. Highly purified steviol glycosides began to be widely used by the public after they were recognized as generally safe in the United States in 2008. Also, on November 11, 2011, the European Food Safety Authority approved the use of purified steviol glycosides in food products equivalent to 4 mg of steviol per kilogram of human body weight per day [6]. Extracts from the Stevia plant are used directly in the form of powders and tablets to produce various sweeteners. The consumption of stevia has increased significantly in the production of pickled vegetables, dried seafood, soy sauces and meat products [7, 8]. In addition, stevia leaves and steviol glycosides are used in the production of carbonated and non-carbonated soft drinks, teas, yogurts, pastries and ice cream. They are also a sweet flavor additive in toothpastes, mouthwashes, and chewing gums [6, 9-12].

The growing demand for natural ingredients with sweetening and antioxidant properties is the main reason why the stevia market is growing in popularity. Increasing use of stevia in the pharmaceutical industry is also fueling its growth. According to data, the stevia market was valued at \$490.1 million in 2017, and will reach \$771.5 million by 2022 [13]. Stevia leaf extract, powder, stevioside and rebaudioside A of various degrees of purity, depending on the type of product and its characteristics, the international prices of sweeteners vary from 10 to 150 US dollars per kg [14]. China is currently the world's largest producer and supplier of stevia sweeteners, but the industry has seen significant growth worldwide in recent years.

The biochemical composition of stevia leaves, rich in natural components, has been found to have beneficial effects on human health [15]. Some of the beneficial effects of stevia consumption have been reported in a number of scientific articles.

Stevia extract has been shown to have beneficial effects on blood glucose levels [12, 16 - 21]. A review article reported that steviol glycosides contained in stevia extract control the

production of insulin and glucagon hormones in the blood [22], have a hypoglycemic effect by reducing the release of glucagon and increasing the release of glucose in the urine.



To date, more than 40 steviol glycosides have been identified from the composition of the stevia plant, which differ only in the number and type of monosaccharides attached to the aglycone steviol ring in the R1 and R2 states. At the C-13 and C-19 positions, the monosaccharides are glucose, rhamnose, xylose, fructose, and deoxyglucose through the 1,2-oxygen; 1,3-; 1,4- or 1,6- α - or β -glycosidic bonds are attached [27-29].

The extraction of steviol glycosides was shown to be significantly less in hexane, chloroform, and ethyl acetate fractions, and 60.3% in the isobutanol fraction. Methanol, ethanol, and ethyl acetate extracts from stevia plant leaves were more rich in phenolic compounds with antioxidant activity than the water extract. According to research data, organic compounds and mineral elements obtained from stevia leaves constitute the main composition [10].

Research methods

The leaf of the plant "Stevia - *Stevia rebaudiana* Bertoni (Asteraceae family)" grown in Uzbekistan was used as a source for obtaining the sweetened purified dry extract in laboratory conditions. Stevia plant leaf grown in Uzbekistan was used as a source to obtain the sweetened purified dry extract in laboratory conditions.

Extraction using water. To obtain stevia dry extract samples, 2.0 kg of the above-ground part of the plant was crushed (fineness level 0.3-0.5 cm) and extracted with distilled water (in a ratio of 1:10) at 70-80 °C (repeated 5 times).

When the composition of the aqueous extract was analyzed by the YUQX (Thin-Layer chromatography) method, it was found that it consists of a small amount of diterpene glycosides (stevioside, rebaudioside A, rebaudioside B, rebaudioside C, etc.) along with water-soluble phenolic

compounds, monosaccharides and other additives. .

The condensed extract was dried to a powder in a desiccator vacuum cabinet and a 1:1 ratio of dis. when diluted in water and treated with organic solvents - chloroform, n-butanol, fractions with the following mass were obtained.

Butanol fraction. After washing the aqueous extract with chloroform and ethyl acetate, it was washed with n-butanol. As a result, a collection of sweet-tasting compounds consisting of 7.55% of dry steviol glycosides in relation to the raw mass was isolated.

Fraction with chloroform - 19.7 g (0.985%);

Ethyl acetate fraction - 20.5 g (1.05%);

Butanol fraction - 150 g (7.55%).

Fertigplatten Kieselgel 60 (Merck) plates were used for thin-layer chromatography, with a suitable chloroform:methanol:water (60:30:6) system selected. A 20x10 cm long plate was placed in the chamber until it rose to the front line in the system and was removed from the chamber and dried at room temperature. A 50% aqueous solution of H₂SO₄ was used to remove spots on the plate.

Stevioside R_f = 0.44, water solubility 0.13%, rebaudioside A R_f = 0.40, water solubility 0.80%, rebaudioside B R_f = 0.52, water solubility 0.10%, rebaudioside C R_f = 0.17, water solubility is 0.80%.

In conclusion, the aqueous extract of the Stevia plant leaf was obtained, for chemical analysis 19.7 g (0.985%) with chloroform, 20.5 g (1.05%) with ethyl acetate, 150 g (7, 55%) were allocated.

Analytical method in high performance liquid chromatography.

High-performance liquid chromatography "Agilent 1200", complete with Degasser G1379A degasser, QuatPump G1311A pump, ALS G1313A autosampler, Colcom G1316A thermostatic column, data processing Agilent ChemStation Rev system with refractometric RID G1362Ai detector developed in USA B.01.03. SupelcosiLLC-NH2 5micron 4.6x250 mm "Supelco" column. "VWR" Poland 100 and 1000 µl micropipette. 5 ml pipette, "Biohit" Finland. AnD GR-202 (accuracy of 0.00001 g) analytical balance, "AnD" Japan. Millipore Simplicity, a water deionizer from Millipore, France. C 30 H

Elmasonic "Elma" German ultrasonic bath. 0.45 micron 13 mm nylon filter. Standards Fructose, Glucose, Sucrose, Maltose (Sigma-aldrich USA). Acetonitrile for YuSSX "Sigma-aldrich" USA.

A method for determining the antimicrobial activity of aqueous, alcoholic extract and chloroform fraction isolated from stevia leaves against typical strains of conditionally pathogenic microorganisms. Antimicrobial activity of the oil isolated from chestnut seeds against typical strains of conditionally pathogenic microorganisms was determined according to the methodical manual OFS.1.2.4.0010.15 "Method for determination of antimicrobial activity by agar diffusion method".

Test microorganisms: conditional pathogenic strains of Escherichia coli NS 101, Pseudomonas aeruginosa 003841/114, Proteus mirabilis 6, Bacillus subtilis VKM, Klebsiella oxitoca 1, Staphylococcus aureus D2, Candida albicans were used as test microorganisms.

The cups were kept at +4 °C for 24 hours so that the extracts and fractions were well absorbed into the agar. Then it was grown at a temperature of 36±1 °C for 16-18 hours. After 18 hours, using a ruler, the diameter of the zones where the growth of test microorganisms was stopped was measured and recorded in a notebook.

Results and Discussion

When the composition of the aqueous extract was analyzed by the YUQX method, it was determined by the YUSSX method that, along with water-soluble monosaccharides, phenolic compounds and other additives, steviol glycosides consist of stevioside, rebaudioside A, rebaudioside B, and rebaudioside C.

When performing high performance liquid chromatography analysis, the standard solution was prepared by accurately weighing it during development. To ensure long-term storage, the standard solution is mixed with acetonitrile in a ratio of 1/1. Working solutions of standards were prepared by diluting the standard solution with a 1/1 water-acetonitrile mixture.

During the development of the method, the analysis conditions were determined: - isocratic elution mode, the composition of the mobile phase is acetonitrile/water in volume 82/18, two separate vessels without mixing. The composition

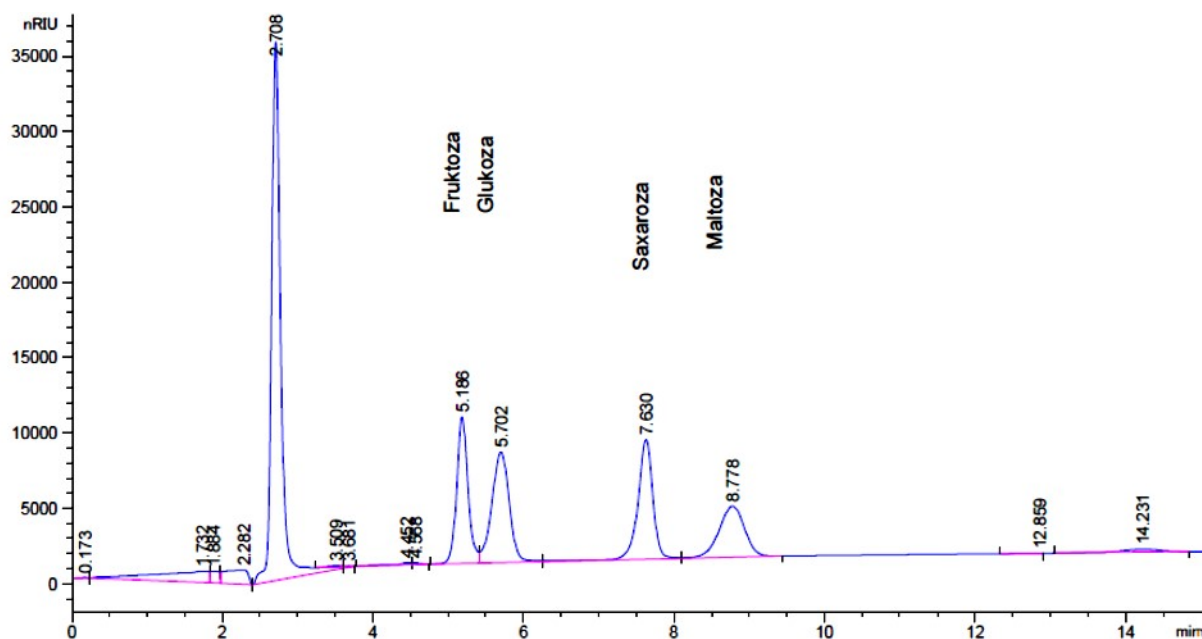


Figure 1. Derived diagram of standard monosaccharides YuSSX.

of the mobile phase can be varied to achieve complete separation of glucose and fructose peaks. Volume speed of the washer is 1.0 ml/min.; injection volume 10 µl; the temperature of the column thermostat is 35 °C; retention times of standards: fructose – 5.18±0.2 min, glucose – 5.70±0.2 min, su-

crose – 7.63±0.2 min, maltose – 8.77±0.2 min. The detector signal of each carbohydrate content was determined in 0.1 concentration range. 10.0 mg/ml at 6 points (corresponding to 10-1000 g/kg extract), repeated 3 times for each point (Table 1).

In the analysis of the aqueous extract of the

Indicators of standard monosaccharides obtained in YuSSX

Table 1

Peak#	RetTime [min]	Type	Width [min]	Field [nRIU*s]	Height [nRIU]	Field %
1	0.173	BB	0.1200	297.81693	39.66695	0.0378
2	1.732	BV	0.6254	3.79169e4	754.19446	4.8171
3	1.884	VV	0.1161	6407.14844	772.96533	0.8140
4	2.282	VB	0.2821	2.00485e4	936.66089	2.5470
5	2.708	BV R	0.1271	3.03288e5	3.57517e4	38.5304
6	3.509	VV E	0.2457	1022.83710	65.19008	0.1299
7	3.681	VB X	0.0811	258.67029	48.93864	0.0329
8	4.452	BV	0.2055	1713.75867	111.33444	0.2177
9	4.568	VB	0.1057	578.38495	80.18887	0.0735
10	5.186	BV	0.1583	1.01248e5	9715.90625	12.8628
11	5.702	VV	0.2507	1.18126e5	7328.91797	15.0071
12	7.630	VV	0.2092	1.09680e5	7912.44922	13.9340
13	8.778	VB	0.3647	8.02477e4	3390.02026	10.1949
14	12.859	BB	0.4043	117.37947	3.49894	0.0149
15	14.231	BV R	0.4455	6187.49658	182.37737	0.7861

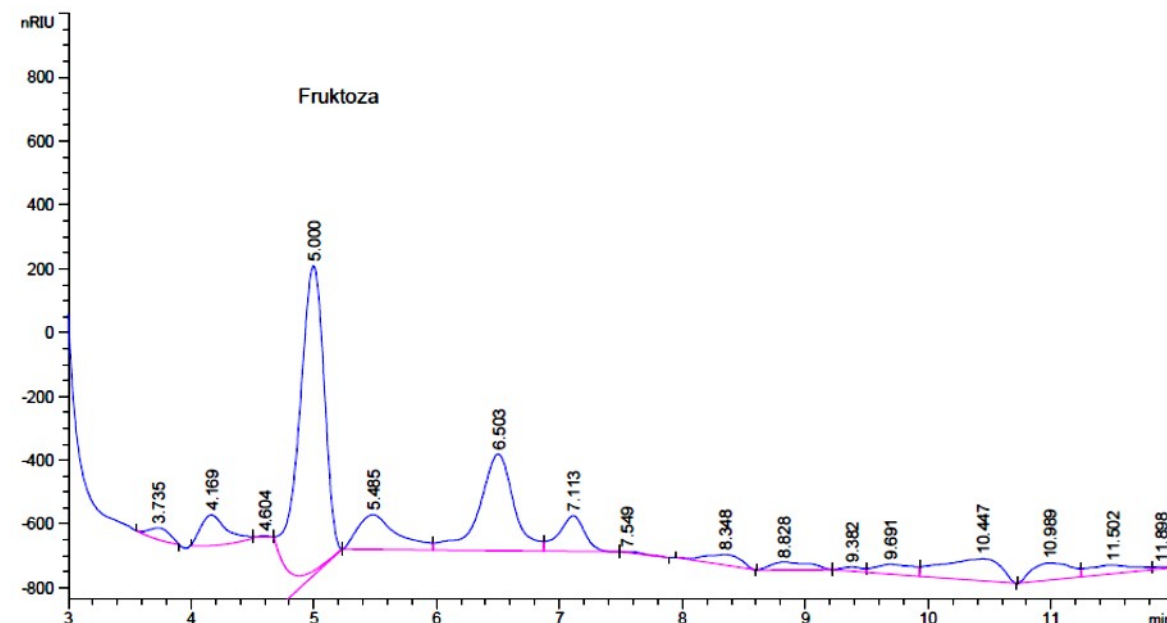


Figure 2. Schematic diagram of aqueous extract of stevia leaf.

stevia plant, the solutions were prepared as follows: 1 g of the studied (exactly weighed) dried extract of stevia was dissolved in 50 ml of deionized water. The solution is mixed in an ultrasonic bath at room temperature in the "mixing" mode until the extract is completely homogeneous. Then the solution is filtered on a membrane filter with a diameter of 0.45 microns. The filtrate was mixed with acetonitrile in a ratio of 1/1 and analyzed using YuSSX (Fig. 2).

Volume speed of the washer is 1.0 ml/min.; injection volume 10 μ l; the temperature of the column thermostat is 35 $^{\circ}$ C; retention times of standards: fructose – 5.00 \pm 0.2 min, glucose – 5.48 \pm 0.2 min, sucrose – 7.11 \pm 0.2 min, maltose – 8.34 \pm 0.2 min. The detector signal of each carbohydrate content was determined in 0.1 concentration range. 10.0 mg/ml at 6 points (corresponding to 10-1000 g/kg extract), repeated 3 times for each point (Table 2).

Antimicrobial activity of aqueous, alcoholic extract and chloroform fraction isolated from stevia leaves against typical strains of conditionally pathogenic microorganisms.

During the experiment, the antimicrobial activity of the studied substances - the aqueous, ethanolic extract of stevia leaves, and the chloroform fraction showed a moderate level of antimicrobial activity against all 7 tested microorganisms. During the experiment, the diameter of the zone of stopping the growth of the test micro-

organisms tested with ethanol extract of stevia: *Proteus mirabilis* 6 – 18 mm, *Pseudomonas aeruginosa* 003841/114 – 15 mm, *Staphylococcus aureus* D2 – 15 mm, *Candida albicans* – 18 mm, *Escherichia coli* NC 101 - 18 mm and *Bacillus subtilis* VKM - 18 mm. This substance did not show antimicrobial activity against *Klebsiella oxitoca* test strain.

Aqueous extract of stevia showed antagonistic activity against 3 tested test microorganisms, and the diameter of their growth arrest zone was: *Pseudomonas aeruginosa* 003841/114 - 12 mm, *Candida albicans* - 19 mm, *Klebsiella oxitoca* - 15 mm. This substance did not show antimicrobial activity against the test strains of *Proteus mirabilis* 6, *Staphylococcus aureus* D2, *Escherichia coli* NC 101 and *Bacillus subtilis* VKM (Table 1).

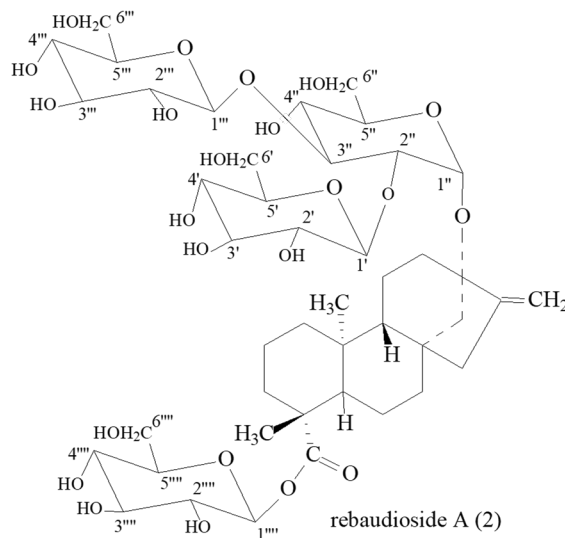
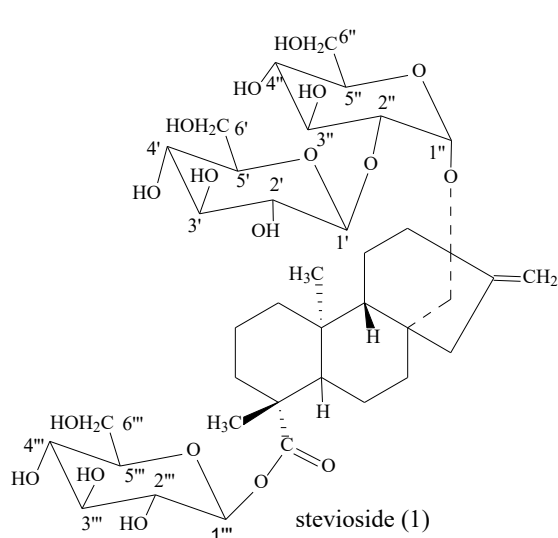
During the experiment, the chloroform fraction showed antagonistic activity against 4 of the tested test microorganisms, and the diameter of their growth arrest zone was: *Proteus mirabilis* 6 - 15 mm, *Staphylococcus aureus* D2 - 13 mm, *Candida albicans* - 12 mm, *Escherichia coli* NC 101 - was 15 mm. This substance did not show antimicrobial activity against *Pseudomonas aeruginosa* 003841/114, *Escherichia coli* NC 101 and *Bacillus subtilis* VKM test strains.

From the results presented in Table 3, it is known that the gram-positive, spore-forming rod-shaped bacterium - *Bacillus subtilis* BKM is sen-

Table 2

YuSSX indicators of aqueous extract of stevia leaves

Peak	RetTime [min]	Type	Width [min]	Field [nRIU*s]	Height [nRIU]	Field %
1	0,025	BV	0,2804	139,88625	8,31485	8,810e-3
2	0,153	VB	0,0829	202,65739	40,73582	0,0128
3	0,583	BB	0,3341	536,96326	20,18726	0,0338
4	0,957	BV	0,1550	619,00653	55,94121	0,0390
5	1,055	VB	0,1163	340,93765	47,42666	0,0215
6	1,425	BV	0,2255	3038,43262	187,81677	0,1914
7	1,717	VV	0,2766	1,00019e4	490,91568	0,6299
8	2,289	VB	0,3049	2,21052e4	938,94025	1,3922
9	2,678	BV R	0,1592	1,51929e6	1,55691e5	95,6850
10	3,735	VV E	0,1983	445,76321	37,14242	0,0281
11	4,169	VV E	0,2221	1407,73315	96,44479	0,0887
12	4,604	VV X	0,0750	18,64134	3,07648	1,174e-3
13	5.000	VB X	0.2185	1.34676e4	955.30316	0.8482
14	5,485	BV	0,3218	2428,89063	108,44365	0,1530
15	6,503	VV	0,2933	5874,00195	302,72379	0,3699
16	7,113	VB	0,2133	1625,62854	111,47227	0,1024
17	7,549	BB	0,1817	54,01439	3,64062	3,402e-3
18	8,348	VB R	0,2772	678,30914	33,20749	0,0427
19	8,828	BB	0,2835	549,66370	24,90006	0,0346
20	9,382	BV	0,1481	143,34172	13,21794	9,028e-3
21	9,691	VV	0,2836	661,51904	30,45890	0,0417
22	10,447	VB	0,4476	2250,69067	68,98055	0,1417
23	10,989	BV	0,2871	1159,75586	53,58004	0,0730
24	11,502	VV	0,3236	715,26611	26,68050	0,0450
25	11.898	VBA	0.1104	48.66902	5.96125	3.065e-3



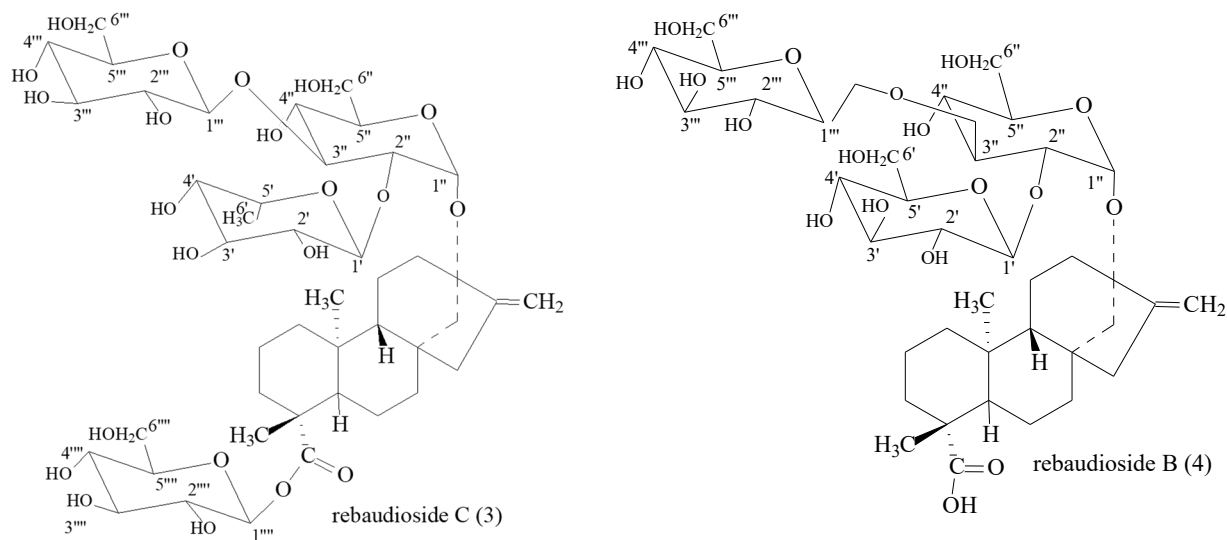


Figure 3. Steviol glycosides present in *Stevia rebaudiana* leaves.

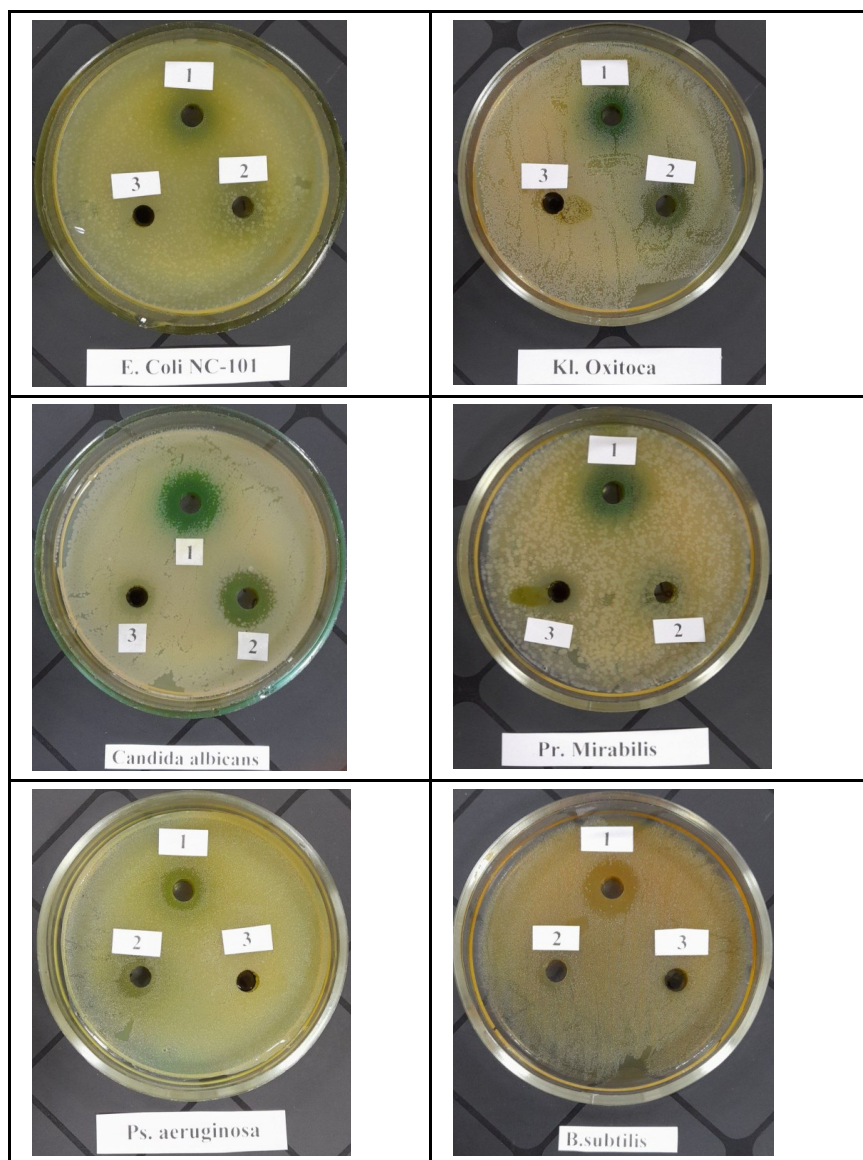


Figure 4. Antimicrobial activity of stevia leaf extracts and fra against opportunistic pathogenic microorganisms. 1. Ethanol extract; 2. Aqueous extract; 3. Fraction with chloroform.

Table 3

Antimicrobial activity of stevia leaf extracts and fraction against conditionally pathogenic microorganisms

№	Conditionally pathogenic microorganism strains	Diameter of the zone of antimicrobial activity of the tested substance, mm		
		Ethanol extract	Aqueous extract	Chloroform fraction
1	Escherichia coli NC 101	18	0	15
2	Proteus mirabilis 6	18	0	15
3	Pseudomonas aeruginosa 003841/114	15	12	0
4	Klebsiella oxitoca 1	0	15	0
5	Staphylococcus aureus D2	15	0	13
6	Bacillus subtilis BKM	18	0	0
7	Candida albicans	18	19	12

sitive to the ethanolic extract of stevia, and other substances do not have an antimicrobial effect on this test microorganism. Also, among the substances tested, only the aqueous extract of stevia showed activity against gram-negative, rod-shaped bacteria *Klebsiella oxitoca* 1. All tested compounds show the same level of antagonistic activity against the yeast *Candida albicans* (Fig. 3).

Conclusion

An aqueous extract of stevia plant leaf was taken, and for chemical analysis, chloroform 19.7 g (0.985%), ethylacetate 20.5 g (1.05%), and n-butanol fraction were divided into 150 g (7.55%).

Using high-performance liquid chromatog-

raphy, monosaccharides in aqueous extract of stevia plant were compared with standard samples.

Chemical analysis of alcoholic and aqueous extracts and chloroform fraction from the stevia plant grown in Uzbekistan was carried out, as a result of which their biochemical composition was determined. In the obtained analysis, the analysis of the main compounds of the sweetener used in the food industry, water-soluble compounds, monosaccharides, steviol glycosides and other natural substances is carried out using YuSSX. Antimicrobial activity of aqueous and alcoholic extracts and fractions obtained from *Styvia* plant was studied, as a result, it was found that the aqueous extract has anti-*Candida albicans* activity.

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